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Chromosomal regions underlying noncoagulation of milk in Finnish Ayrshire cows

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Motivation to study milk coagulation properties ?

- Milk coagulation is a critical step in cheese-making
- 40% of milk is used for cheese production in Finland
- Milk coagulation properties are impaired in Finnish Ayrshire (Tyrisevä 2008)







Coagulation time R (min) Curd-firming time K₂₀ (min) Curd firmness E₃₀ (mm) Genetic correlations between the above traits are almost one





Definitions for noncoagulating and coagulating

If E₃₀ = 0 mm, milk is noncoagulating (NC)

If E₃₀ < 20 mm poorly coagulating</p>

If $E_{30} > 30$ mm excellently coagulating (E)



About 12% of Finnish Holstein-Friesian and about 30% Finnish Ayrshire cows produce poorly coagulating milk (Tyrisevä et al. (2004))
 About 1% of Finnish Holstein-Friesian and about 10% Finnish Ayrshire cows produce noncoagulating milk (Ikonen et al. (1999), Tyrisevä et al. (2003), Tyrisevä et al. (2004))



If excellently coagulating, poorly coagulating and noncoagulating milk are mixed, the mixture has impaired coagulation properties → both quality and yield of cheese are impaired



Heritability estimates (with SE)
 Milk coagulation time R (min) 0.28 (0.03)
 Curd firmness E₃₀ (mm) 0.39 (0.04)
 Coefficients of repeatability
 Milk coagulation time R (min) 0.66
 Curd firmness E₃₀ (mm) 0.68



To locate chromosomal regions affecting noncoagulation of milk in Finnish Ayrshire



Studied traits:

curd firmness (E₃₀)

somatic cell score

■ pH of milk

Two stages:

(1) genome scan

(2) verification of the genome scan



Genome scan

Genome scan with 194 microsatellite markers using selective DNA pooling method (Darvasi and Soller 1994) Pooled DNA samples

33 NC-milk producing cows

49 E-milk producing cows

Allele intensities were tested for homogeneity between the two pools using the method of Sham and Curtis (1995)



Verification of the genome scan

16 regions on 11 chromosomes were selected
 47 microsatellite markers

18 sires and 477 daughters individually genotyped

188 NC-milk producing cows

289 E-milk producing cows

- Maximum likelihood method (ANIMAP, Georges et al.1995)
- Nonparametric method (HSQM, Coppieters et al. 1998)



Sire families were a sub-sample from Ikonen et al. (2004)
 Only sires assumed to be heterozygous for the hypothesized NC-genes and sires with large daughter groups were selected for genome scan





Based on daughters' distribution of milk coagulation ability, sires can be classified to homozygous (A), heterozygous (B) and non-carriers for the NCgenes (C)





In genome scan, 16 markers in 11 chromosomes reached the 1% statistical significance threshold
 In verification study, the most significant findings on chromosomes 2, 18 and 24 for noncoagulation
 Putative findings for pH and milk coagulation ability on chromosomes 24 and 27



Mapping results across families

Chr	Marker	-log(p)	Experimentwise risk level
2	BMS1126	4.3	0.001
18	BMS1355	4.3	0.001



Interval mapping using nonparametric method (2)

Mapping results on chromosomes 24 and 27

Chr	Trait	Family	Max	-log(p)	Experimentwise	
			position		risk level	
24	NC	38651	BM7151	4.3	<0.001	
24	рН	38585	AGLA269	2.4	<0.05	
27	NC	38585	DIK5134	2.9	<0.05	
27	рН	38651	DIK5134	3.7	<0.01	



Interval mapping using maximum likelihood

Estimates for effects

Family	Chr	Marker position	LOD	Experimentwise risk level	Effect (phenotypic SD)
38651	18	BMS1355	2.7	0.05	1.9
38651	24	BM7151	4.2	0.01	1.8



5. Conclusions (1)

 Regions on chromosomes 2 and 18 affect noncoagulation of milk
 Putative findings

 chromosomes 18 and 24 associated with noncoagulation of milk in single families
 markers on chromosomes 24 and 27 associated with coagulation ability and pH of milk



5. Conclusions (2)

Further mapping needed on chromosomes 2 and 18 before marker assisted selection is viable option to improve milk coagulation ability by elimination of carrier bulls of NC-genes

Dissection of candidate chromosomal regions is ongoing using SNP markers



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