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**Genetic polymorphism at *CSN1S1*, *CSN2* and *CSN3* loci in the Holstein and Czech Fleckvieh breeds kept in the Czech Republic.**

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***Abstract***

The dairy industry continuously strives to improve the quality of its products. It has long been known that manufacturing properties of milk are influenced by the relative composition of its proteins. The bovine casein cluster contains four milk protein genes, which are closely linked, in the order *CSN1S1*, *CSN2*, *CSN1S2* and *CSN3*. Several polymorphisms have been described for each casein gene. The objective of this work was to determine genetic polymorphism in the candidate genes *CSN1S1*, *CSN2* and *CSN3* in Holstein and Czech Fleckvieh breeds kept in the Czech Republic. The casein genes from a 184 Holstein (H) and 502 Czech Fleckvieh (CF) were determined by PCR-RFLP methods and LightCycler analysis. Results showed that at the locus *CSN1S1*, allele *B* was predominant in both breeds (0.965 and 0.852). Allele *A2* was predominant in both breeds (0.818 and 0.644) at the *CSN2* locus, with allele *A* predominant (0.788 and 0.640, respectively) at the locus *CSN3*.

***Introduction***

Casein proteins are the major constituents (80%) of total protein in bovine milk. These proteins have been extensively studied, and it has been reported that specific genetic variants of the caseins affect cheese making properties (Grosclaude et al., 1988; Martin et al., 2002). Significant associations with several milk production traits have also been identified for sites of polymorphism within noncoding regions in the casein complex (Martin et al., 2002). The casein loci have been physically mapped within a 200-kb fragment on the bovine chromosome 6 (*BTA6q31-33*) (Treadgill and Womack, 1990) in the order *CSN1S1-CSN2-CSN1S2-CSN3*. The loci are considered to influence milk production traits (Bovenhuis et al., 1992). Antibacterial activities of derived peptides (Malkoski et al., 2001) may also affect the biological fitness of the offspring.

Casein genes could thus be subjected to natural and artificial selection (Ward et al., 1997). Genetic polymorphisms in the casein genes allow the determination of casein haplotypes. The latter can be used for studies concerning quantitative traits (Lien et al., 1995; Velmala et al., 1995) or phylogeny (Mahe et al., 1999), since they provide more information than individual genes (Lien and Rogne, 1993).

Most of the casein alleles have been identified at the protein level and differ from each other by a few base substitutions that cause one or two amino acid changes in the protein. The methods, which are used to detect genetic variation on DNA level are based on PCR and its

modifications e.g. PCR-RFLP, AS-PCR, PCR-SSCP. Further techniques include the hybridization probes (Hoffmann et al., 2007) and microarray technology or ligation detection reaction (LDR) combined with a universal array (UA) (Chessa et al, 2007).

This study aimed at examining the frequency of genotypes and alleles determining individual milk protein fractions (*CSN1S1*, *CSN2* and *CSN3*) in Holstein and Czech Fleckvieh, cattle breeds in the Czech Republic.

### **Material and Methods**

Blood samples were collected from 686 animals (184 Holstein and 502 Czech Fleckvieh) for genomic DNA extraction using ABI PRISM 6100 analysis (Nucleic acid Prep. Station, Applied Biosystem, Co) according to the standard protocol. DNA fragments of the three casein genes were amplified by standard PCR technology.

Genetics variants *B* and *C* at the *alpha* S1 (*CSN1S1*) gene was typed for *Mae*III polymorphism in the promoter region (*CSN1S1prom*) by PCR-RFLP according to Koczan et al. (1993) with some modifications (primer sequence). *Beta* casein (*CSN2*) genetic variants *A1*, *A2*, *A3* and *B* were genotyped by PCR and using the LightCycler analysis (unpublished data), whereas *kappa* casein (*CSN3*) gene was genotyped by PCR-RFLP according to the protocol (Schlieben et al., 1991; Lien and Rogne, 1993). In the latter the genetic variants: *A*, *B* and *E* have been evaluated.

### **Statistical analysis**

Allele and genotype distribution, Hardy-Weinberg equilibrium, polymorphic information contents and heterozygosity were estimated using PowerMarker software V3.25 (Liu and Muse, 2005).

### **Results**

Results of the molecular analysis of the bovine *CSN1S1*, *CSN2* and *CSN3* casein genes are presented in Table 1 and 2. Litter difference in the occurrence and frequencies of different alleles and genotypes at the casein loci were found between in Holstein and Fleckvieh breeds kept in the Czech Republic.

The *B* and *C* variants of the *CSN1S1* casein gene were observed in both Holstein and Czech Fleckvieh breeds with the *B* variant predominant in both breeds (Table 1). The *CSN1S1* *CC* genotype was absent and the heterozygote genotype *BC* showed a low frequency only in the Holstein population. In the Czech Fleckvieh breed genotype *BC* and *CC* presented a low frequencies with the highest frequency found for genotype *BB*. According to Lien and Rogne (1993); Jann et al., (2004), and Chessa et al. (2007) the superiority of the frequency of alleles *CSN1S1 B* and *CSN1S1 C* is characteristic for almost all breeds of European cattle.

In both breeds, the genetic variant *A2* at the *CSN2* casein gene was predominant, followed by *A1*. Allele *A3* and *B* showed a very low frequency in both breeds (Table 1). Genotype frequencies were observed in the order *A2A2* > *A1A2* > *A1A1* > *A2A3* > *A1A3* > *A2B*, *A3B* in the Czech Fleckvieh breed. In the Holstein breed, the order was *A1A2*, *A2A2* > *A1A1* > *A2B* > *A1B* > *A2A3*. Genotypes *A1A3*, *A3A3*, *A3B* and *BB* in the Czech Fleckvieh, and *A1B*, *A3A3* and *BB* in the Holstein population were absent (Table2).

At the *CSN3* casein gene, variant *A*, was predominant in both breeds, with different trends depending on breed and origin. Allele *B* and *E* showed low frequencies in both breeds (Table 1). In the Czech Fleckvieh population, *AA* and *AB* were found to be the predominant genotype compared to genotypes *AE*, *BB* and *BE*. Genotype *EE* was absent in both cattle

populations. In Holsteins, genotypes *AE*, *BB* and *BE* were observed at low frequencies. Genotype *AB* and *AE* had a medium frequency, and genotype *BB* and *BE* showed very low frequencies in the Holstein population.

Large similarities regarding the casein loci, *CSN1S1*, *CSN2* and *CSN3* have been reported among Holstein populations (Poli and Antonini, 1991) and in some European bovine breeds (Jann et al., 2004).

All loci were tested for the Hardy-Weinberg equilibrium and characterised by expected ( $H_{exp}$ ) heterozygosity and polymorphic information content (PIC) in both cattle populations. The Czech Fleckvieh population followed the Hardy-Weinberg equilibrium at all evaluated loci. However, the Holstein population did not follow the Hardy-Weinberg equilibrium the *CSN2* locus. The smallest heterozygosity was observed at the *CSN1S1* in the Holstein breed and the highest was described at the *CSN3* in the Czech Fleckvieh breed. PIC showed a very low value at the *CSN1S1* locus in Holsteins, with the highest value observed at the *CSN3* locus in the Czech Fleckviehs.

### **Conclusion**

Allele and genotype distribution at all loci in Holstein and the Czech Fleckvieh agree with results previously described in northern, central European and southern European cattle breeds as well as in Holstein populations. Differences in allele distribution depend on origin, intensity of selection and domestication history among cattle breeds.

Although further studies are needed to evaluate the haplotype distribution on the both Holstein and the Czech Fleckvieh breeds and effect on milk protein production.

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### **References**

- Bovenhuis, H., van Arendonk, J. a., Korver, S.** (1991) Association between milk protein polymorphisms and milk production traits. *J. Dairy Sci.* 75, 2549-2559.
- Chessa, S., Chiatti, F., Ceriotti, G., Caroli, A., Consolandi, C., Pagnacco, G., Castiglioni, B.** (2007) Development of a single nucleotide polymorphism genotyping microarray platform for the identification of bovine milk protein genetic polymorphisms. *J. Dairy Sci.* 90, 451-464.
- Grosclaude F.** (1988) Le polymorphisme génétique des principes lactoprotéines bovine. *INRA Production Animales* 1, 5-17
- Hoffmann, M., Hurlebaus, J., Weilke, Ch.** (2007) Novel methods for high-performance melting curve analysis using the LightCycler 480 system. In: *Biochemica* 1, 2007, Roche Applied Science, pp 17-19.
- Jann, O. C., Ibeagha-Awemu, E. M., Özbeyaz, C., zaragoza, P., Williams J. L., Ajmone-Marsan, P., Lenstra, J. A., Moazami-Goudarzi, K., Erhardt, G.** (2004) Geographic distribution of haplotype diversity at the bovine casein locus. *Genet. Sel. Evol.* 36, 243-257.
- Lien, S., Gomez-Raya, L., Teine T., Fimland, E., Rogne, S.** (1995) Association between casein haplotypes and milk yield traits. *J. Dairy Sci.* 78, 2047-2056.
- Lien, S., Rogne, S.** (1993) Bovine casein haplotypes: number, frequencies and applicability as genetic markers. *Anim. Genet.* 24, 373-376.

- Liu, K. and Muse, S.V.** (2005) PowerMarker: Integrated analysis environment for genetic marker data. *Bioinformatics* 21(9): 2128-2129.
- Malkoski, M., Dasher S. G., O'Brien-Simpson, N. M., Talbo, G. H., Macris, M., Cross, K. J., Reynolds, E. C.** (2001) Kappacin, a novel antibacterialpeptide from bovie milj. *Antimicrob. Agents Chemother.* 45, 2309-2315.
- Martin P., Szymanowska M., Zwierzchowski L., Leroux Ch.** (2002): The impact of genetic polymorphisms on the protein composition of ruminant milks. *Reprod. Nutr. Dev.* 42, 433-459.
- Poli, M. A., Antonini, A. G.** (1991) Genetic structure of milk proteins in Argentinian Holstein and Argentinian Creole cattle. *Hereditas* 115, 177-182.
- Schlieben, S., Erhardt, G., Senft, B.** (1991) Genotyping of bovine  $\kappa$ -casein ( $\kappa$ -CN<sup>A</sup>,  $\kappa$ -CN<sup>B</sup>,  $\kappa$ -CN<sup>C</sup>,  $\kappa$ -CN<sup>E</sup>) following DNA sequence amplification and direct sequencing of the  $\kappa$ -CN<sup>E</sup> PCR product. *Anim. Genet.* 22, 333-342
- Threadgill D. V.,Womack J. E.** (1990) Genomic analysis of the major bovine – milk protein genes. *Nucleic Acid Res.* 18, 23, 6935-6942.
- Velmala, R., Vilkki, J., Elo, K., Maki-Tanila, A.** (1995) Casein haplotypes and their association with milk production traits in the Finnish Ayrshire cattle. *Anim. Genet.* 26, 419-425.
- Ward, T. J., Honeycutt, R. L., Derr, J. N.** (1997) Nucleotide sequence evolution at the kappa-casein locus: evidence for positive selection within the family Bovidae. *Genetics* 147, 163-1872.

**Table 1** Allele distributions, heterozygosity and PIC of the *CSN1S1*, *CSN2* and *CSN3* loci in Holstein and Fleckvieh cattle breed.

Locus	Czech Fleckvieh			Holstein			
	Frequency	Heterozygosity	PIC	<i>CSN1S1</i>	Frequency	Heterozygosity	PIC
<i>CSN1S1</i>							
<i>B</i>	0.852			<i>B</i>	0.964		
<i>C</i>	0.148	0.245	0.221	<i>C</i>	0.036	0.07	0.066
<i>CSN2</i>				<i>CSN2</i>			
<i>A1</i>	0.328			<i>A1</i>	0.155		
<i>A2</i>	0.644			<i>A2</i>	0.818		
<i>A3</i>	0.001			<i>A3</i>	0.022		
<i>B</i>	0.027	0.472	0.387	<i>B</i>	0.005	0.289	0.274
<i>CSN3</i>				<i>CSN3</i>			
<i>A</i>	0.64			<i>A</i>	0.788		
<i>B</i>	0.326			<i>B</i>	0.106		
<i>E</i>	0.034	0.509	0.394	<i>E</i>	0.106	0.375	0.328

**Table 2** Genotype distributions of the *CSN1S1*, *CSN2* and *CSN3* loci in Holstein and Czech Fleckvieh cattle breed.

Locus	Genotype	Czech Fleckvieh		Holstein	
		No.	Frequency	No.	Frequency
<i>CSN1S1</i>	<i>BB</i>	366	0.729	171	0.929
	<i>BC</i>	123	0.245	13	0.071
	<i>CC</i>	13	0.026	-	-
<i>CSN2</i>	<i>A1A1</i>	56	0.112	5	0.027
	<i>A1A2</i>	209	0.416	44	0.24
	<i>A1A3</i>	-	-	3	0.016
	<i>A1B</i>	8	0.016	-	-
	<i>A2A2</i>	209	0.416	126	0.685
	<i>A2A3</i>	1	0.002	4	0.022
	<i>A2B</i>	19	0.038	1	0.005
	<i>A3A3</i>	-	-	-	-
	<i>A3B</i>	-	-	1	0.005
	<i>BB</i>	-	-	-	-
<i>CSN3</i>	<i>AA</i>	201	0.400	113	0.614
	<i>AB</i>	222	0.442	30	0.163
	<i>AE</i>	19	0.038	34	0.185
	<i>BB</i>	45	0.090	2	0.011
	<i>BE</i>	15	0.030	5	0.027
	<i>EE</i>	-	-	-	-