

Isolation of antibacterial peptides from rabbit milk casein

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Introduction

Milk proteins have been considered for a long time for their nutritional value only, namely to supply the organism with essential amino acids. It is well established that proteolytical digestions of bovine and human caseins yield several bioactive peptide fragments, which are involved in the regulation of various physiological processes. Peptides derived from casein hydrolysates show immuno-modulatory and anti-inflammation effects, play important roles in passive immunity, are important for the growth and the development, inhibit platelet aggregation, and alter gastrointestinal function. Therefore milk proteins are important factors not only in consideration of their nutritional value, but also for their intrinsic potentiality to be used as dietary supplements or in future as drugs [for reviews see 1, 2, 3].

Objectives

Caseins represent a heterogeneous class of milk proteins whose bactericidal properties have been poorly investigated. Some antibacterial activity has been ascertained for some peptides derived from the bovine casein only. The aim of our work was to search whether bactericidal peptides could be generated from the digestion of rabbit casein by endopeptidase present in the gastrointestinal tract. We expect from one side to learn more about the physiological function of casein and on the other side to explore the possibility to obtain antimicrobial compounds from food proteins.

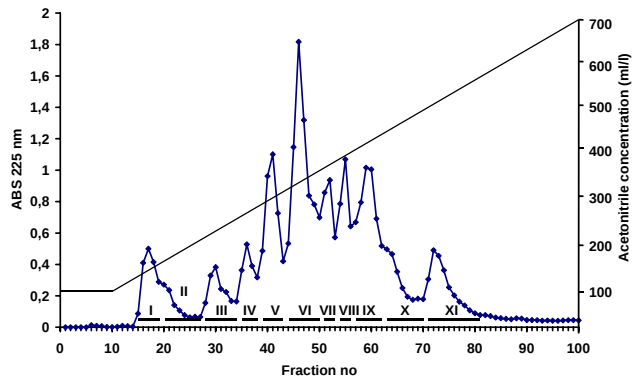
Isolation of peptides

whole rabbit casein

enzyme (trypsin)

incubation at 37°C

RP-FPLC



Collection of fractions: Fr. 1, 2, 3, 4, 5, 6, 7, 8, 100 (200)

pooling fractions belonging to the different peaks

Pool I, II, III, IV, V,

FREEZE-DRYING

RP-FPLC

Pools with antibacterial activity

antibacterial assay

Isolated, antibacterial peptide

antibacterial assay

RP-FPLC

purification

peptide synthesis

sequencing

1. step (column 120-10)

Pool I	Pool II	Pool III	Pool IV	Pool V	Pool VI	Pool VII	Pool VIII	Pool IX	Pool X	Pool XI
E.coli -	E.coli -	E.coli -	E.coli -	E.coli -	E.coli -	E.coli -	E.coli -	E.coli -	E.coli -	E.coli 0,5
B.subtilis 0.81	B.subtilis 1.77	B.subtilis 0.40	B.subtilis 0.03	B.subtilis 1.12	B.subtilis -	B.subtilis -	B.subtilis -	B.subtilis -	B.subtilis 1.42	B.subtilis >3.0
St. lentus 0.17	St. lentus 0.08	St. lentus -	St. lentus -	St. lentus 0.37	St. lentus 0.49	St. lentus -	St. lentus 0.04	St. lentus 0.04	St. lentus 0.95	St. lentus 0.10

2. step (column 100-5)

Pool 1	Pool 2
B.subtilis 0.62	B.subtilis 0.75
St. lentus -	St. lentus -

2. step (120-10 oszlop)

Pool 1
B.subtilis 0.44
St. lentus 0.07

2. step (column 120-10)

Pool 1
B.subtilis 0.44
St. lentus 0.07

2. step (column 120-10)

Pool 1	Pool 2
B.subtilis 0.15	B.subtilis -
St. lentus -	St. lentus -

Mixed by a mistake

2. step (column 120-10)

Pool 1	Pool 2	Pool 3	Pool 4
B.subtilis 0.18	B.subtilis 0.04	B.subtilis -	B.subtilis 0.12
St. lentus 0.06	St. lentus -	St. lentus 0.08	St. lentus -

2. step (column 120-10)

Pool 1	Pool 2	Pool 3
B.subtilis 0.44	B.subtilis 0.23	B.subtilis 0.60
St. lentus 0.35	St. lentus 0.25	St. lentus 0.12

3. step (column 100-5)

Pool 1	Pool 2
B.subtilis -	B.subtilis -
St. lentus -	St. lentus -

3. step (column 120-10)

Pool1	Pool 2	Pool 3	Pool 4
St. lentus 1.00	St. lentus 0.46	St. lentus 0.31	B.subtilis 0.56
			St. lentus 0.23

3. step (column 100-5)

Pool 1	Pool 2
B.subtilis -	B.subtilis -
St. lentus -	St. lentus -

3. step (column 100-5)

Pool1
B.subtilis 2.12
St. lentus -

sequenced!
TP-3 (ILPFIQSLFPFAER)

4. step (column 100-5)

Pool 1
St. lentus 0.46

sequenced!
TP-1 (HVEQLLR)

4. step (column 100-5)

Pool 1	Pool 2
St. lentus -	St. lentus -

4. step (column 100-5)

Pool 1	Pool 2
St. lentus 0.13	St. lentus 0.09

4. step (column 100-5)

Pool 1
St. lentus -

5. step (column 100-5)

Pool 1
St. lentus 0.11

sequenced!
TP-2 (FHLGHLK)

5. step (column 100-5)

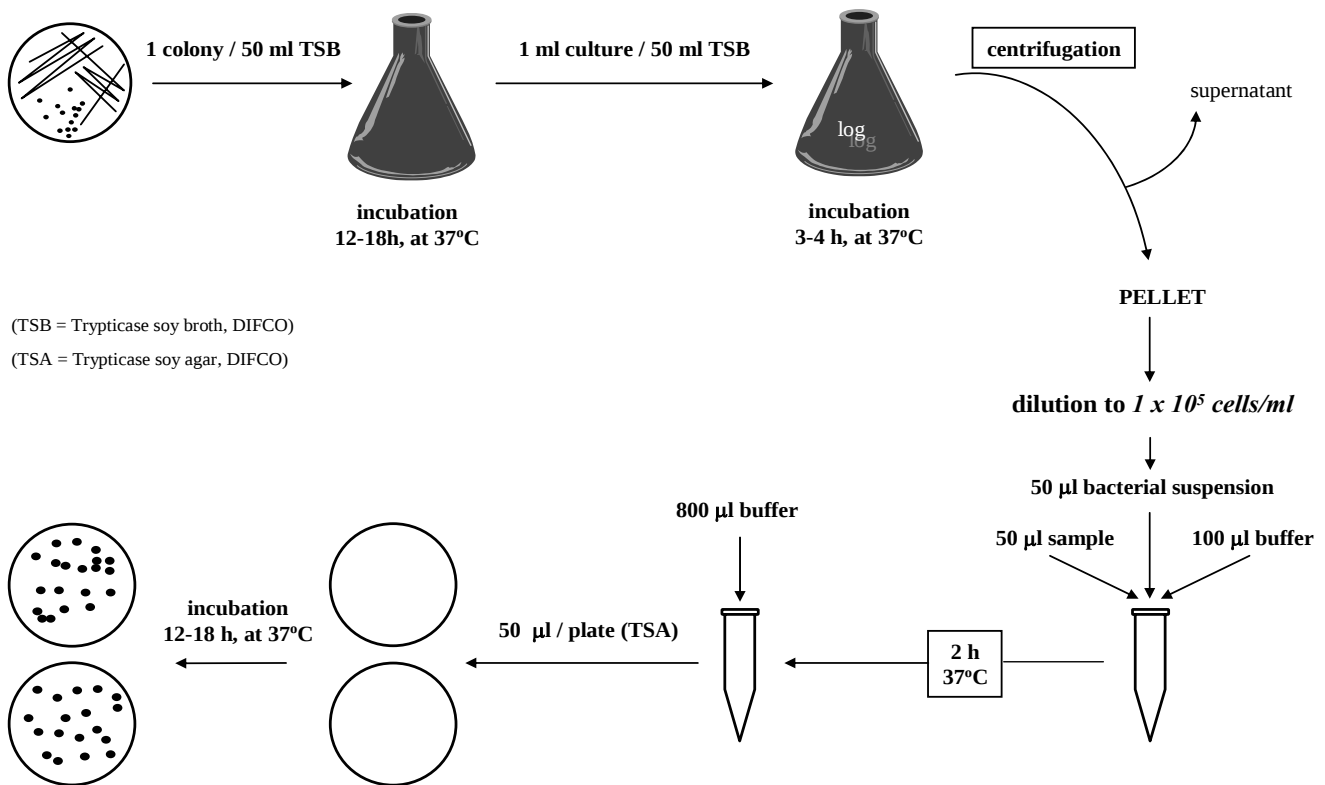
Pool 1	Pool 2
St. lentus 0.11	St. lentus 0.09

6. step (column HD)

Pool 1	Pool 2
St. lentus 0.06	St. lentus -

Chromatographic separation

Antibacterial assay



(TSB = Trypticase soy broth, DIFCO)

(TSA = Trypticase soy agar, DIFCO)

Calculation of antibacterial activity

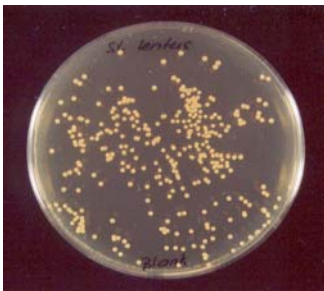
$$\log (N_0 / N_1)$$

$N_0 =$

number of control colonies

$N_1 =$

number of colonies grown in the presence of antibacterial agent



Testing the pools

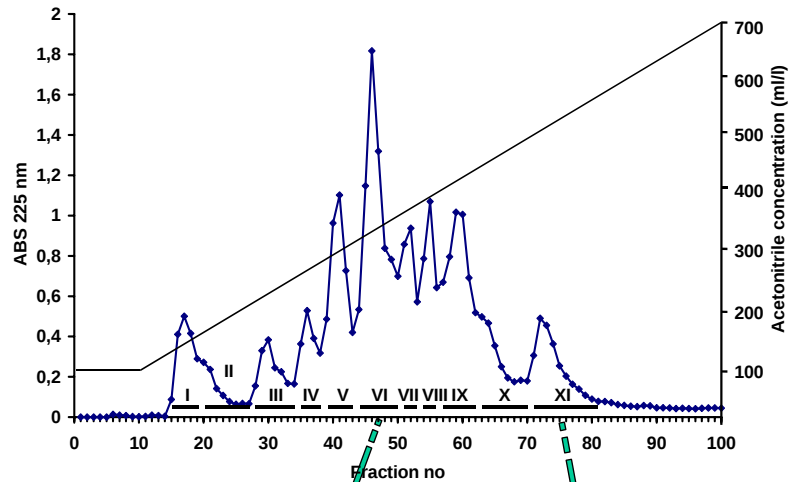
Bacteria	Strain	Type
<i>Escherichia coli</i>	ATCC 25922	Gram-neg.
<i>Bacillus subtilis</i>	BGA	Gram-pos.
<i>Staphylococcus lentus</i>	WS	Gram-pos.
<i>Klebsiella pneumoniae</i>	ATCC 13883	Gram-neg.
<i>Serratia marcescens</i>	ATCC 8100	Gram-neg.
<i>Streptococcus zooepidemicus</i>	WS	Gram-pos.

Testing synthetic peptides

Results

Trypsin digestion

	<i>Esch. coli</i>	<i>B. subtilis</i>	<i>Staph. lentus</i>
Pool I	0	0.81 ± 0.37	0.17 ± 0.02
Pool II	0	1.77 ± 0.26	0.08 ± 0.02
Pool III	0	0.40 ± 0.03	0
Pool IV	0	0.03 ± 0.03	0
Pool V	0	1.12 ± 0.12	0.37 ± 0.07
Pool VI	0	0	0.49 ± 0.13
Pool VII	0	0	0
Pool VIII	0	0	0.04 ± 0.02
Pool IX	0	0	0.04 ± 0.02
Pool X	0	1.42 ± 0.09	0.95 ± 0.10
Pool XI	0.50 ± 0.30	>3.00 ± 0.00	0.10 ± 0.02



Further purification of pools VI and XI

HVEQLLR
(beta-casein)

FHLGHLK
(alpha-S1-casein)

ILPFIQSLFPFAER
(beta-casein)

Rabbit casein-derived antibacterial peptides

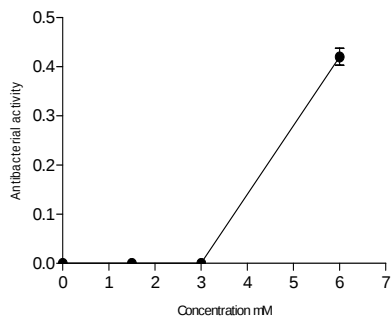
<i>Rabbit alpha-S1-casein</i>				
10	20	30	40	50
MKLLILTCLV	ATLARHK FH TP-2 LGHLK LKTQEQ	PESSEQEILK	ERKLLRFVQT	
60	70	80	90	100
VPLELREEYV	NELNRQRELL	REKENEEIKG	TRNEVTEEHV	LADRETEASI
110	120	130	140	150
SSSSEEVPS	STKQKYVPRE	DLAYQPYVQQ	QLLRMKERYO	IQEREPMRVV
160	170	180	190	200
NQELAQLYLQ	PFEQPYQLDA	YLPAPWYYTP	EVMQVYVLSPL	FYDLVTPSAF
210	215			
ESAEKTDVIP	EWLKN			

<i>Rabbit beta-casein</i>				
10	20	30	40	50
MKVLILACL	ALALAREKEQ	LSVPTEAVGS	VSSSEEITHI	NKQKLETIK H
60	70	80	90	100
TP-1 VEQLL REEKL	QDK ILPFIQS TP-3 LFPFAER IPY	PTLPQNILNL	AQLDMLLPLL	
110	120	130	140	150
QPEIMEDPKA	KETIIPKHKL	MPFLKSPKTV	PFVDSQILNL	REMNQHL
160	170	180	190	200
PQLLPFMHQV	FQFPFQTPIP	YPQALLSLPQ	SKFMPIVPQV	VPYPQRDMPI
210	220	228		
QALQLFQELL	FPTHQGYPVV	QPIAPVNV		

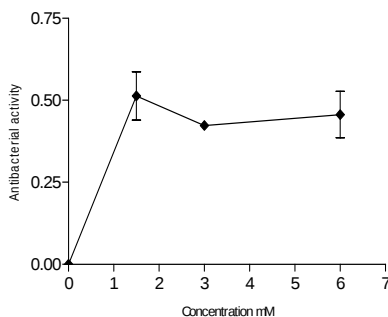
Peptide	sequence	casein	position	pI
TP-1	HVEQLLR	β-CN	50-56	6,5
TP-2	FHLGHLK	α _{S1} -CN	19-25	8,7
TP-3	ILPFIQSLFPFAER	β-CN	64-77	6,0

Antibacterial activity of synthetic peptides

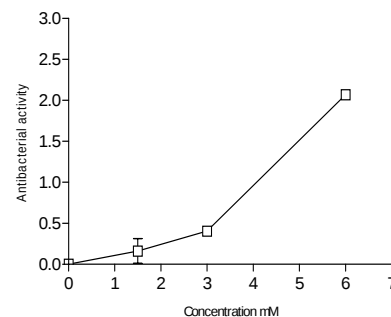
B. subtilis



**HVEQLLR
(TP-1)**

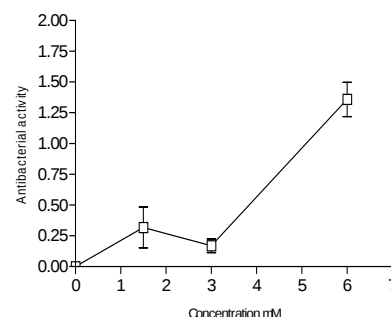
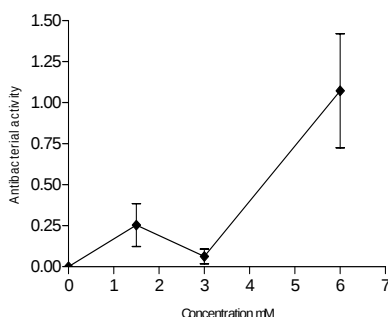
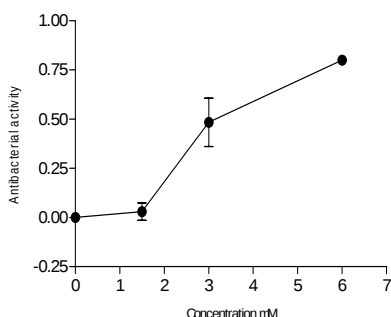


**FHLGHLK
(TP-2)**



**ILPFIQSLFPFAER
(TP-3)**

Staph. lentus



Summary

Whole rabbit casein was digested with trypsin and the arising peptide fragments were separated by reversed-phase FPLC. According to their amino acid sequence the identified peptides were synthesized and the antibacterial activity of the synthetic peptides tested against six Gram positive and Gram negative bacterial strains.

★ **Trypsin digestion:**

- 😊 4 pools strongly, 1 pool moderately inhibited the growth of *B. subtilis*
- 😊 2 pools moderately inhibited the growth of *Staph. lentus*

★ **Pools with antibacterial activity were further purified and the amino acid sequence of the isolated peptides was determined:**

- 😊 2 antibacterial peptides were identified from pool VI
HVEQLLR (beta-casein, residues 50-56, pI 6.5) and
FHLGHLK (alpha-S1-casein, residues 19-25, pI 8.7)
- 😊 1 antibacterial peptide was identified from pool XI
ILPFIQSLFPFAER (beta-casein, residues 64-77, pI 6.0).

★ **Synthetic peptides:**

- 😊 Active only against the Gram-positive bacteria *B. subtilis* and *Staph. lentus*, but not against *Strep. zooepidemicus*. Not active against Gram-negative bacteria *Esch. coli*, *Kl. pneumoniae*, and *Ser. marcescens*.

Acknowledgements

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References

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- 2) Schanbacher, F.L., Talhouk, R.S. and Murray, F.A., Livestock Production Science 50 (1997) 105.
- 1) Meisel, H., Livestock Production Science 50 (1997) 125.